

PROTOCOL

Ferret Igκ Single-Color ELISPOT Assay

CONTENTS:

- Anti-ferret Igκ Primary Ab
- Anti-ferret Igκ (Biotin) Secondary Ab
- SA-AP
- S1 (Substrate component 1)
- S2 (Substrate component 2)
- S3 (Substrate component 3)
- Diluent B (2 bottles)
- Diluent C
- Diluent Blue
- Plates: 96-well, high-protein-binding, PVDF filter plates
- Protocol

CRITICAL REAGENTS NOT INCLUDED:

- Coating antigen
- Low-binding 0.1µm syringe filters

STIMULATE B CELLS

- Cells need to already possess antibody-secreting capability directly ex vivo prior to isolation.

COAT THE ELISPOT PLATE

- Prepare antigen coating solution in PBS (coating conditions should be optimized for each test antigen to achieve maximal assay sensitivity) and prepare 70% EtOH (see Solutions).
- **Pre-wetting:** Remove underdrain from the PVDF plate and set aside. Pipette 15µl of 70% EtOH into each well and then immediately add 180µl/well of PBS. Decant and wash again with 180µl/well of PBS.
Note: Activation of the membrane with EtOH is instantaneous and can be seen visually as a graying of the membrane. It is important to be sure that the EtOH solution has wicked across the entire membrane of all wells before adding the first PBS wash.
- **Coating:** Decant PBS from plate, blot excess on paper towel and replace the underdrain. Do not press directly on the bottom of the underdrain. Instead, line up the underdrain, and put pressure on the plate frame to spread force evenly across the entire plate. Quickly pipette 80µl/well of the antigen coating solution.
- Seal plate in parafilm or place in a humid chamber and incubate overnight at 4°C.

ELISPOT Protocol

CELL HARVEST AND PLATING

- Isolate cells and adjust to desired concentration using the assay medium.
- **Washing and blocking:** Decant the antigen coating solution plated the previous day.
- Add 150µl/well of sterile PBS to remove excess unbound antibody or antigen.
- Decant PBS, blot excess on paper towel and add 150µl/well of assay medium.
- Incubate for a minimum of 1 hour at room temperature.
- Decant plate and add 100µl/well of assay medium.
- Add 100µl/well of assay medium containing cells at the desired concentration using large orifice tips. Once completed, gently tap the sides of the plate to distribute cells and immediately place into a 37°C humidified incubator, 5-10% CO₂.
- Incubate for 4-8 hours at 37°C. Do not stack plates. Avoid shaking plates by carefully opening and closing incubator door. Do not touch plates during assay incubation.

DETECTION

- **Prepare wash solutions:** Prepare Wash Buffers: PBS, distilled water and PBS-T (see Solutions).
- **Detection:** Prepare Anti-ferret Igκ Primary Solution (see Solutions).
- After the completion of assay incubation, decant assay medium containing cells and wash plate twice with 150µl/well of PBS.
- Wash plate twice with 150µl/well of PBS-T. Decant final wash and blot excess on paper towel without allowing the membrane to dry.
- Add 80µl/well of Anti-ferret Igκ Primary Solution. Incubate for a minimum of 1 hour in the dark at room temperature or overnight at 4°C.

For laboratory research use only. Not for use in diagnostic or therapeutic procedures.



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DETECTION CONTINUED

- Prepare Anti-ferret Igκ Secondary Solution (See Solutions).
- Wash plate twice with 150µl/well of PBS-T. Decant final wash and blot excess on paper towel without allowing the membrane to dry.
- Add 80µl/well of Anti-ferret Igκ Secondary Solution. Incubate for 1 hour in the dark at room temperature.
- Prepare Tertiary Solution (See Solutions).
- Wash plate twice with 150µl/well of PBS-T. Decant final wash and blot excess on paper towel without allowing the membrane to dry.
- Add 80µl/well of Tertiary Solution. Incubate for 30 minutes in the dark at room temperature.
- Prepare CTL-TrueBlue Substrate Solution (see Solutions).
- Decant and wash plate twice with 150µl/well of distilled water. Decant final wash and blot excess on paper towel without allowing the membrane to dry.
- Add 80µl/well of CTL-TrueBlue Substrate Solution. Incubate for 10 minutes in the dark at room temperature.
- Decant plate and remove underdrain. Wash both sides of the plate gently using tap water. Decant final wash and blot excess on paper towel.
- Dry plate in a running laminar flow hood or overnight face down on paper towel in the dark.
- Scan and count plate. CTL has scanning and analysis services available. Email kitscanningservices@immunospot.com to inquire.

SOLUTIONS

- All solutions should be freshly-made prior to use. It is important to quick-spin vials contained in the kit before use to ensure content volumes.
- 70% EtOH: Dilute 190-200 proof EtOH. For 10ml, add 7ml of EtOH to 3ml of distilled water.
- Assay medium: Prepare assay medium (see Technical Tips) by adding fresh L-glutamine to a final concentration of 2mM. The amount of medium needed will depend on variables such as cell yield and number of samples tested, but will not be less than 35ml for one full plate; warm to 37°C before using.
- Primary Solution: Dilute Anti-ferret Igκ Primary Ab in Diluent B. For one plate, add 10µl of Anti-ferret Igκ Primary Ab to 10ml of Diluent B and filter through a low-protein binding 0.1µm filter (Millipore# SLVVR33RS or SLVVM33RS) (see Technical Tips).
- Secondary Solution: Dilute Anti-ferret Igκ (biotin) Secondary Ab in Diluent B and filter. For one plate, add 10µl of Anti-ferret Igκ (Biotin) Secondary Ab to 10ml of Diluent B and filter through a low-protein binding 0.1µm filter.
- Tertiary Solution: Dilute SA-AP in Diluent C. For one plate, add 10µl of SA-AP to 10ml of Diluent C and filter through a low-protein binding 0.1µm filter.
- Substrate Solution: Dilute substrate components by adding reagents in order. It is recommended to make the Substrate Solution within 10 minutes of use and protect from direct light.
 - CTL-TrueBlue: For one plate:
 - Step 1 – Aliquot 10ml of Diluent Blue.
 - Step 2 – Add 160µl of S1 and mix well.
 - Step 3 – Add 160µl of S2 and mix well.
 - Step 4 – Add 92µl of S3 and mix well.
- Wash Buffers (**not included**): For each plate prepare 200ml distilled water, 200ml PBS and 200ml of PBS-T (100µl Tween-20 into 200ml PBS = 0.05% v/v).

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TECHNICAL TIPS

- Upon successful completion of the assay Igκ spots will appear blue.
- A suitable assay medium for usage in B cell ImmunoSpot[®] is RPMI 1640 with pre-tested 10% FCS, 2mM L-glutamine, 100U/ml Penicillin, 100µg/ml Streptomycin, 8mM HEPES and 50µM 2-mercaptoethanol.
- Deviations from specified temperatures, timing requirements, number of washing steps and specified reagent preparation volumes may alter the performance of the assay. Longer incubation times beyond 8 hours may increase assay sensitivity, but also can increase background membrane staining. Longer substrate incubation times will result in darker spots.
- It is recommended to use a low-protein binding 0.1µm filter for filtering reagents after diluting in the diluents provided. However, if it is not possible to filter, high speed centrifugation of the antibody prior to transferring required volume into the diluents provided, is an alternative.
- The coating concentration of antigens used for ImmunoSpot[®] assays needs to be optimized by the end user. Only pre-wet and coat one to two plates at a time to avoid drying of the membrane before addition of coating solution.
- It is recommended to prepare the next reagent solution prior to decanting the plate to avoid drying of the membrane between detection steps.
- For antigen-specific assays, an initial cell concentration of 1-5 x 10⁵ cells/well may be required. A serial titration of cells is recommended in order to determine the optimal cell concentration. CTL recommends using the CTL-LDC[™] Kit for accurate live/dead cell counts.
- The PVDF membrane is permeable and protected by an underdrain. Do not push the underdrain too far up on the plate after the pre-wetting step, as this may result in leaking of the membrane. Instead, pressing down on the plate frame or the lid itself is preferred. It is OK if the underdrain falls off after the cell incubation, just avoid direct contact of the membrane with your hands, wet surfaces or absorbent materials. To prevent damage to the PVDF membrane in the bottom of the well, avoid touching with pipette tips or the plate washer.
- The PVDF membrane must remain wet during the detection steps. After blotting the plate on paper towel to remove excess liquid, immediately add the next detection solution to avoid the membrane from drying.
- Scan and count plates only after they have completely dried. ELISPOT plates are stable and can be scanned weeks to months after assay completion without any loss of signal when stored away from direct light.
- Optimal removal of fibers and debris is achieved by removing the underdrain, flipping the plate face down and filtering distilled water "back to front" through the plate using a vacuum manifold (Millipore #MSVMHTSOO).
- Plates may be washed manually or using a suitable plate washer with adjusted pin length and flow rate so membranes and spots are not damaged (CTL recommends the CTL 405LSR).
- Data analysis: The CTL ImmunoSpot[®] Analyzers along with the ImmunoSpot[®] Software have advanced features that permit automated, objective recognition of spots, gating and counting. An ELISPOT data management tool, SpotMap[®], is also available to facilitate high-throughput ELISPOT work.
- CTL has scanning and analysis services available. Email kitscanningservices@immunospot.com to inquire. Please consult with Technical Support for assistance at +1-216-791-5084.

The CTL team can assist with data analysis and troubleshooting, along with customizing ImmunoSpot[®] assays to suit your needs. Please contact us at kits@immunospot.com

Visit our YouTube channel for several helpful videos on working with ELISPOT assays and PBMC:

www.youtube.com/user/ImmunoSpot.

The CTL Thawing Protocol for cryopreserved human PBMC is available at www.immunospot.com.



PROTOCOL: Ferret Igκ Single-Color ELISPOT Assay

B cell ImmunoSpot® Assay

When provided with the appropriate stimuli in vivo, B cells acquire a lymphoblastoid/plasmablast phenotype and begin to secrete their unique B cell receptor as immunoglobulin (Ig); more commonly referred to as antibody. Such Ig-secreting plasmablasts, also known as antibody-secreting cells (ASC), can be directly plated and measured in B cell ImmunoSpot® assays. This system enables measurement of B cells secreting Igκ antibodies directly ex vivo.

Principle of the Antigen-Specific Test

The principle for detection of antigen-specific Igκ ASC is illustrated in the figure below. The membrane is initially coated with the antigen itself and only ASC producing antibody with sufficient binding strength for the plate-bound antigen will generate a secretory footprint (shown in grey). These antigen-specific Igκ secretory footprints are then detected by adding an Anti-ferret Igκ Primary Ab (shown in light blue), followed by a biotinylated Anti-ferret Igκ Secondary Ab (shown in dark blue). Finally, addition of Streptavidin-Alkaline Phosphatase (SA-AP) (shown in red) and then a substrate that consists of three subcomponents (S1, S2 and S3); which generates a blue precipitate or "spots".

